Code:	0198
Cell Line:	P388
Species:	Mus musculus **Vulgar Name:** Mouse; Dba/2
Morphology:	Macrophage
Disease:	Lymphoid Neoplasm
Growth Properties:	Adherent
Derivation:	Originating in a DBA/2 mouse as lymphoid neoplasm methylcholantrene-induced, P388 cells was converted to ascit form in the first mouse transfer.
Biosafey:	1
Additional info:	The in vitro derived line as obtained from 49 consecutive mouse passages. These cells actively phagoc
Culture Medium:	RPMI 1640 with 2.0 mM L-glutamine adjusted to contain 1.5 g/L sodium bicarbonate, 4.5 g/L glucose, 10.0 mM HEPES, and 1.0 mM sodium pyruvate, 90%; fetal bovine serum, 10%.
Subculturing:	Subcultures are prepared by scraping. Remove old medium, add fresh dislodge the cells and dispense into new flasks. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010. **Medium Renewal:** 3 times per week **Subcultivation ratio:** 1:4 to 1:8
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)
Thawing Frozen	SAFETY PRECAUTION: Is highly recommend that protective gloves

Cells:	and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4.Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in a appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).
References:	Amer. J. Pathol. 3: 33, 1957.
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