

## Data Sheet

PAGE 1/3

<b>BCRJ Code:</b>	0001
<b>Cell Line:</b>	11B11 [BSF-1]
<b>Species:</b>	Rattus norvegicus (B cell); Mus musculus (myeloma), rat (B cell); mouse (myeloma)
<b>Vulgar Name:</b>	Mouse / Rat
<b>Cell Type:</b>	Hybridoma: B Lymphocyte
<b>Morphology:</b>	Lymphoblast
<b>Growth Properties:</b>	Suspension
<b>Derivation:</b>	Animals were immunized with purified BSF-1 emulsified in complete Freund's adjuvant. Spleen cells were fused with P3-NSI/1-Ag4-1 myeloma cells.
<b>Applications:</b>	The antibody does not cross-react with other lymphokines, and can be used to purify BSF-1 from complex mixtures.
<b>Products:</b>	immunoglobulin; monoclonal antibody; against murine B cell stimulatory factor 1 (BSF-1, interleukin-4, interleukin 4, IL-4, B cell growth factor 1, BCGF-1)
<b>Biosafety:</b>	1
<b>Culture Medium:</b>	RPMI 1640 medium with L-glutamine and 10% of heat-inactivated fetal bovine serum.
<b>Subculturing:</b>	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at $1 \times 10^5$ viable cells/mL. Maintain cultures at a cell concentration between $1 \times 10^5$ and $1 \times 10^6$ cells/mL. NOTE: Do not allow the cell concentration to exceed $1 \times 10^6$ cells/mL.
<b>Subculturing Medium Renewal:</b>	Every 2 to 3 days



## Data Sheet

PAGE 2/3

**Culture Conditions:** Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

**Cryopreservation:** 95% FBS + 5% DMSO (Dimethyl sulfoxide)

**SAFETY PRECAUTION:** It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).

2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.

3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes.

4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).

5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

**NOTE:** It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

## Thawing Frozen Cells:

## References:

Ohara J, Paul WE. Production of a monoclonal antibody to and molecular characterization of B-cell stimulating factor-1. *Nature* 315: 333-336, 1985. PubMed: 2582266 Hay, R. J., Caputo, J. L., and Macy, M. L., Eds. (1992), ATCC Quality Control Methods for Cell Lines. 2nd edition, Published by ATCC. Caputo, J. L., Biosafety procedures in cell culture. *J. Tissue Culture Methods* 11:223-227, 1988. Fleming, D.O., Richardson, J. H., Tulis, J.J. and Vesley, D., (1995) *Laboratory Safety: Principles and Practice*. Second edition, ASM press, Washington, DC.

**Depositors:** Ises A. Abrahamsohn, Universidade de São Paulo



**Cellosaurus:**

[CVCL\\_9139](#)