

## Data Sheet

<b>BCRJ Code:</b>	0001
<b>Cell Line:</b>	11B11 [BSF-1]
<b>Species:</b>	Rattus norvegicus (B cell); Mus musculus (myeloma), rat (B cell); mouse (myeloma)
<b>Vulgar Name:</b>	Mouse / Rat
<b>Cell Type:</b>	Hybridoma: B Lymphocyte
<b>Morphology:</b>	Lymphoblast
<b>Growth Properties:</b>	Suspension
<b>Derivation:</b>	Animals were immunized with purified BSF-1 emulsified in complete Freund's adjuvant. Spleen cells were fused with P3-NSI/1-Ag4-1 myeloma cells.
<b>Applications:</b>	The antibody does not cross-react with other lymphokines, and can be used to purify BSF-1 from complex mixtures.
<b>Products:</b>	immunoglobulin; monoclonal antibody; against murine B cell stimulatory factor 1 (BSF-1, interleukin-4, interleukin 4, IL-4, B cell growth factor 1, BCGF-1)
<b>Biosafety:</b>	1
<b>Culture Medium:</b>	RPMI 1640 medium with L-glutamine and 10% of heat-inactivated fetal bovine serum.
<b>Subculturing:</b>	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at $1 \times 10^5$ viable cells/mL. Maintain cultures at a cell concentration between $1 \times 10^5$ and $1 \times 10^6$ cells/mL. NOTE: Do not allow the cell concentration to exceed $1 \times 10^6$ cells/mL.
<b>Subculturing Medium Renewal:</b>	Every 2 to 3 days

## Data Sheet

PAGE 2/2

**Culture Conditions:** Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

**Cryopreservation:** 95% FBS + 5% DMSO (Dimethyl sulfoxide)

### Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. It is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). **NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

### References:

Ohara J, Paul WE. Production of a monoclonal antibody to and molecular characterization of B-cell stimulating factor-1. *Nature* 315: 333-336, 1985. PubMed: 2582266 Hay, R. J., Caputo, J. L., and Macy, M. L., Eds. (1992), ATCC Quality Control Methods for Cell Lines. 2nd edition, Published by ATCC. Caputo, J. L., Biosafety procedures in cell culture. *J. Tissue Culture Methods* 11:223-227, 1988. Fleming, D.O., Richardson, J. H., Tulis, J.J. and Vesley, D., (1995) *Laboratory Safety: Principles and Practice*. Second edition, ASM press, Washington, DC.

**Depositors:** Ises A. Abrahamsohn, Universidade de São Paulo

**ATCC:** HB-188