

# Banco de Células do Rio de Janeiro

### **Data Sheet**

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**BCRJ Code:** 0002

Cell Line: 145-2C11

Cricetulus migratorius (B cell); Mus musculus (myeloma), hamster, Armenian (B **Species:** 

cell); mouse (myeloma)

**Vulgar Name:** Hamster / Mouse

Tissue: Blood

Cell Type: Hybridoma: B Lymphocyte

Morphology: Lymphoblast-Like

**Growth Properties:** Suspension

Animals were immunized with the BM10-37 mouse cytotoxic T lymphocyte **Derivation:** 

(CTL) cell clone (anti H-2 Kb). Spleen cells were fused with Sp2/0-Ag14 myeloma

cells.

It reacts with all mature T cells and can both activate and inhibit T cell function.

The antibody is specific for a 25000 dalton protein component (CD3 epsilon) of the antigen specific T cell receptor. The antibody reacts with the murine T cell receptor (CD3 - T3) complex. The antibody does not react with peripheral blood lymphocytes from rats, rabbits, miniature swine or hamsters. It reacts with all

mature T cells and can both activate and inhibit T cell function.

**Products:** immunoglobulin; monoclonal antibody; against mouse CD3

Biosafety: 1

**Applications:** 

The origin of this cell line should be acknowledged in all relevant publications. **Addtional Info:** 

May be distributed to scientific institutions; not to be distributed for any comm

DMEM with 4 mM L-glutamine, 4500 mg/L glucose and 10% of fetal bovine **Culture Medium:** 

serum.

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## **Subculturing:**

Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1 x 10e5 viable cells/mL. Maintain cultures at a cell concentration between 1 x 10e5 and 1 x 10e6 cells/mL. NOTE: Do not allow the cell concentration to exceed 1 x 10e6 cells/mL.

# Subculturing Medium Renewal:

Every 2 to 3 days

**Culture Conditions:** 

Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C

### **Cryopreservation:**

95% FBS + 5% DMSO (Dimethyl sulfoxide)

It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4.Discard the supernatant and Resuspend cell pellet with the recommended complete

SAFETY PRECAUTION: Is highly recommend that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials.

#### **Thawing Frozen Cells:**

Leo O, et al. Identification of a monoclonal antibody specific for a murine T3 polypeptide. Proc. Natl. Acad. Sci. USA 84: 1374-1378, 1987. PubMed: 2950524 Kayagaki N, et al. Polymorphism of murine Fas ligand that affects the biological activity. Proc. Natl. Acad. Sci. USA 94: 3914-3919, 1997. PubMed: 9108079

medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in a appropriate atmosphere and

avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15

temperature (see "Culture Conditions" for this cell line). NOTE: It is important to

# References:





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minutes to allow the medium to reach its normal pH (7.0 to 7.6).



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**Depositors:** Alberto Nobrega, Universidade Federal do Rio de Janeiro

**ATCC:** CRL-1975

**Cellosaurus: CVCL 7234** 



