Data Sheet

BCRJ Code:	0002
Cell Line:	145-2C11
Species:	Cricetulus migratorius (B cell); Mus musculus (myeloma), hamster, Armenian (B cell); mouse (myeloma)
Vulgar Name:	Hamster / Mouse
Tissue:	Blood
Cell Type:	Hybridoma: B Lymphocyte
Morphology:	Lymphoblast-Like
Growth Properties:	Suspension
Derivation:	Animals were immunized with the BM10-37 mouse cytotoxic T lymphocyte (CTL) cell clone (anti H-2 Kb). Spleen cells were fused with Sp2/0-Ag14 myeloma cells.
Applications:	It reacts with all mature T cells and can both activate and inhibit T cell function. The antibody is specific for a 25000 dalton protein component (CD3 epsilon) of the antigen specific T cell receptor. The antibody reacts with the murine T cell receptor (CD3 - T3) complex. The antibody does not react with peripheral blood lymphocytes from rats, rabbits, miniature swine or hamsters. It reacts with all mature T cells and can both activate and inhibit T cell function.
Products:	immunoglobulin; monoclonal antibody; against mouse CD3
Biosafety:	1
Addtional Info:	The origin of this cell line should be acknowledged in all relevant publications. May be distributed to scientific institutions; not to be distributed for any comm
Culture Medium:	DMEM with 4 mM L-glutamine, 4500 mg/L glucose and 10% of fetal bovine serum.

Subculturing:	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1 x 10e5 viable cells/mL. Maintain cultures at a cell concentration between 1 x 10e5 and 1 x 10e6 cells/mL. NOTE: Do not allow the cell concentration to exceed 1 x 10e6 cells/mL.
Subculturing Medium Renewal:	Every 2 to 3 days
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)
Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

References:	Leo O, et al. Identification of a monoclonal antibody specific for a murine T3 polypeptide. Proc. Natl. Acad. Sci. USA 84: 1374-1378, 1987. PubMed: 2950524 Kayagaki N, et al. Polymorphism of murine Fas ligand that affects the biological activity. Proc. Natl. Acad. Sci. USA 94: 3914-3919, 1997. PubMed: 9108079
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