

## Data Sheet

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<b>BCRJ Code:</b>	0011
<b>Cell Line:</b>	33D1
<b>Species:</b>	Rattus norvegicus (B cell); Mus musculus (myeloma), rat (B cell); mouse (myeloma)
<b>Vulgar Name:</b>	Rat/Mouse
<b>Cell Type:</b>	Hybridoma: B Lymphocyte
<b>Morphology:</b>	Lymphoblast
<b>Growth Properties:</b>	Suspension
<b>Derivation:</b>	Animals were immunized with mouse spleen and lymph node dendritic cells. Spleen cells were fused with P3X63Ag8-U1 myeloma cells.
<b>Products:</b>	immunoglobulin; monoclonal antibody; against mouse dendritic cells (cytotoxic)
<b>Biosafety:</b>	1
<b>Additional Info:</b>	The antigen recognized by the 33D1 antibody appears to be present only on dendritic cells (10000 to 15000 sites/cell). The antigen does not appear to be immune response associated antigen or Ia antigen.
<b>Culture Medium:</b>	RPMI-1640 medium modified to contain 2 mM L-glutamine, 1 mM sodium pyruvate, 4500 mg/L glucose and fetal bovine serum to a final concentration of 10%.
<b>Subculturing:</b>	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1 x 10 <sup>5</sup> viable cells/mL. Maintain cultures at a cell concentration between 1 x 10 <sup>5</sup> and 1 x 10 <sup>6</sup> cells/mL. NOTE: Do not allow the cell concentration to exceed 1 x 10 <sup>6</sup> cells/mL.
<b>Subculturing Medium Renewal:</b>	Every 2 to 3 days



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**Culture Conditions:** Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

**Cryopreservation:** 95% FBS + 5% DMSO (Dimethyl sulfoxide)

### Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). **NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

### References:

Nussenzweig MC, et al. A monoclonal antibody specific for mouse dendritic cells. Proc. Natl. Acad. Sci. USA 79: 161-165, 1982. PubMed: 6948298 Steinman RM, et al. Dendritic cells are the principal stimulators of the primary mixed leukocyte reaction in mice. J. Exp. Med. 157: 613-627, 1983. PubMed: 6185614

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**ATCC:** TIB-227

