

## Data Sheet

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<b>BCRJ Code:</b>	0042
<b>Cell Line:</b>	B 95-8
<b>Species:</b>	Saguinus oedipus
<b>Vulgar Name:</b>	Monkey - Cotton-Top Tamarin
<b>Tissue:</b>	Peripheral Blood
<b>Cell Type:</b>	Lymphoblast
<b>Morphology:</b>	Lymphoblast/Fibroblast
<b>Growth Properties:</b>	Semiadherent
<b>Derivation:</b>	Derived from a cotton-top tamarin (Saguinus oedipus). Releases high titres of transforming EBV.
<b>Applications:</b>	Thus it provides a source of EBV to establish continuous lymphocytic lines from human donors.
<b>Products:</b>	EBV
<b>Biosafety:</b>	2
<b>Additional Info:</b>	The cells should be handled under laboratory containment level 2. In some instances the cell line B95-8 has been described as derived from 'marmoset', however, this is not correct. B95-8 was derived from a cotton-top tamarin (Saguinas oedipus); this has been confirmed by DNA profiling.
<b>Culture Medium:</b>	RPMI-1640 medium modified to contain 2 mM L-glutamine, 1 mM sodium pyruvate, 4500 mg/L glucose and fetal bovine serum to a final concentration of 10%.

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**Subculturing:**

Subcultures are prepared by diluting the suspension. Cells on the floor of the flask may be dislodged by aspirating several times with culture medium or by rinsing with 0.25% trypsin - 0.53 mM EDTA solution. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.

**Subculturing Medium Renewal:**

Every 2 to 3 days

**Subculturing Subcultivation Ratio:**

1:5 to 1:10 is recommended

**Culture Conditions:**Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C**Cryopreservation:**

95% FBS + 5% DMSO (Dimethyl sulfoxide)

**Thawing Frozen Cells:**

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in a appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

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**References:**

Experientia 1996;52:818-826 - PMID: 8774755 Proc Natl Acad Sci USA  
1972;69:383-387 - PMID: 4333982 Proc Natl Acad Sci USA 1973;70:190-194 -  
PMID: 4346033

**Depositors:**

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**ATCC:**

CRL-1612