

## Data Sheet

PAGE 1/3

<b>BCRJ Code:</b>	0079
<b>Cell Line:</b>	EOMA
<b>Species:</b>	Mus musculus
<b>Vulgar Name:</b>	Mouse/129
<b>Tissue:</b>	Tumor
<b>Cell Type:</b>	Endothelial
<b>Morphology:</b>	Endothelial
<b>Disease:</b>	Hemangioendothelioma
<b>Growth Properties:</b>	Adherent
<b>Age/Ethnicity:</b>	ADULT /
<b>Derivation:</b>	The EOMA cell line was originally derived in 1980 from a mixed hemangioendothelioma arising in an adult mouse.
<b>Tumor Formation::</b>	Yes, in syngeneic mice
<b>Products:</b>	angiotensin converting enzyme (ACE) thrombospondin cathepsin L endostatin interleukin-6 (interleukin 6, IL-6)
<b>Biosafety:</b>	1

## Data Sheet

PAGE 2/3

### Additional Info:

The cells synthesize angiotensin-converting enzyme, express surface receptors for acetylated low density lipoprotein, produce thrombospondin and show intracellular staining with an antibody to von Willebrand factor. Cathepsin L is secreted by EOMA cells and is responsible for the generation of endostatin L. Although constitutive cytokine gene expression exists in EOMA cells, the level of IL-6 mRNA is prominently elevated by incubation with Liposome encapsulated hemoglobin (LEH). The cells constitutively express the vascular addressin identified by antibody MECA-99. EOMA cells exhibit characteristic endothelial cell properties, such as rearrangement into tubelike structures on Matrigel and retention of cobblestone morphology at confluence. They behave in vitro in a manner similar to microvascular endothelial cells.

### Culture Medium:

Dulbecco's Modified Eagle's Medium (DMEM) modified to contain 4 mM L-glutamine, 4500 mg/L glucose, 1 mM sodium pyruvate and fetal bovine serum to a final concentration of 10%.

### Subculturing:

Volumes used in this protocol are for 75 cm<sup>2</sup> flask; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes. T-75 flasks are recommended for subculturing this product. Remove and discard culture medium. Briefly rinse the cell layer with PBS without calcium and magnesium to remove all traces of serum that contains trypsin inhibitor. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.

### Subculturing Medium Renewal:

Every 2 to 3 days

### Subculturing Subcultivation Ratio:

1:3 to 1:6

### Culture Conditions:

Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

### Cryopreservation:

95% FBS + 5% DMSO (Dimethyl sulfoxide)



## Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.
3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes.
4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).
5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

**NOTE:** It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

## References:

47148: Felbor U, et al. Secreted cathepsin L generates endostatin from collagen XVIII. EMBO J. 19: 1187-1194, 2000. PubMed: 10716919 51514: Obeso J, et al. A hemangioendothelioma-derived cell line: its use as a model for the study of endothelial cell biol

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## Cellosaurus:

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