

## Data Sheet

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<b>BCRJ Code:</b>	0086
<b>Cell Line:</b>	FDC-P1
<b>Species:</b>	Mus musculus
<b>Vulgar Name:</b>	Mouse; DbA/2
<b>Tissue:</b>	Bone Marrow
<b>Morphology:</b>	Lymphoblast
<b>Disease:</b>	Normal
<b>Growth Properties:</b>	Suspension
<b>Derivation:</b>	The FDC-P1 cell line was established from long term culture of normal DBA/2 bone marrow cells in medium conditioned by WEHI-3 cells.
<b>Applications:</b>	This cell line is a suitable transfection host.
<b>Biosafety:</b>	1
<b>Additional Info:</b>	The FDCP-1 cell line is dependent upon IL-3 or GM-CSF or WEHI-3 conditioned medium (WEHI-3CM) for continued growth. It can be used to quantify the presence of these growth factors in biological fluids.
<b>Culture Medium:</b>	Dulbecco's modified Eagle's medium with 4 mM L-glutamine with 4.5 g/L glucose, 10% of fetal bovine serum and 25% mouse Interleukin-3 culture supplement. (Supplement purchased from BD Biosciences, Catalog No. 354040).
<b>Subculturing:</b>	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at $1 \times 10^5$ viable cells/mL. Maintain cultures at a cell concentration between $1 \times 10^5$ and $1 \times 10^6$ cells/mL. NOTE: Do not allow the cell concentration to exceed $1 \times 10^6$ cells/mL. Population Doubling Time about: 24-30 hours

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### Subculturing Medium Renewal:

Every 2 to 3 days

### Culture Conditions:

Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

### Cryopreservation:

95% FBS + 5% DMSO (Dimethyl sulfoxide)

### Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.
3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes.
4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).
5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

**NOTE:** It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

### References:

Dexter TM, et al. Growth of factor-dependent hemopoietic precursor cell lines. J. Exp. Med. 152: 1036-1047, 1980. PubMed: 6968334

### Depositors:

Evan secur, USA.

### Cellosaurus:

[CVCL\\_2039](#)



