

## Data Sheet

<b>BCRJ Code:</b>	0327
<b>Cell Line:</b>	GM16000
<b>Species:</b>	Homo sapiens
<b>Vulgar Name:</b>	Human
<b>Tissue:</b>	Blood, Peripheral Vein
<b>Cell Type:</b>	B Lymphocyte
<b>Morphology:</b>	Lymphoblast
<b>Growth Properties:</b>	Suspension
<b>Sex:</b>	Female
<b>Age/Ethnicity:</b>	33 Year / Caucasian
<b>Derivation:</b>	Transformant by Epstein-Barr Virus
<b>Biosafety:</b>	1
<b>Culture Medium:</b>	RPMI-1640 medium modified to contain 2 mM L-glutamine, 4500 mg/L glucose and 10% of fetal bovine serum.
<b>Subculturing:</b>	Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension.
<b>Subculturing Medium Renewal:</b>	Every 2 to 3 days
<b>Subculturing Subcultivation Ratio:</b>	1:3 is recommended

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**Culture Conditions:** Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

**Cryopreservation:** 95% FBS + 5% DMSO (Dimethyl sulfoxide)

### Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the vial and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). **NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

### References:

Bao YP, Huber M, Wei TF, Marla SS, Storhoff JJ, Müller UR, SNP identification in unamplified human genomic DNA with gold nanoparticle probes Nucleic acids research 33:e15 2005 PubMed ID: 15659576 Bernacki SH, Beck JC, Muralidharan K, Schaefer FV, Shrimpton AE, Richie KL, Levin BC, Pont-Kingdon G, Stenzel TT., Characterization of publicly available lymphoblastoid cell lines for disease-associated mutations in 11 genes. Clin Chem 51(11):2156-9 2005 PubMed ID: 16244288 Moser MJ, Marshall DJ, Grenier JK, Kieffer CD, Killeen AA, Ptacin JL, Richmond CS, Roesch EB, Scherrer CW, Sherrill CB, Van Hout CV, Zanton SJ, Prudent JR, Exploiting the enzymatic recognition of an unnatural base pair to develop a universal genetic analysis system. Clin Chem 49(3):407-14 2003 PubMed ID: 12600952

**Depositors:** MARCELO NEVES DE MEDEIROS - INMETRO

**Cellosaurus:** ["CVCL W239 "](#)



