BCRJ Code:	0417
Cell Line:	Grunt Fin (GF)
Species:	Haemulon sciurus
Vulgar Name:	Fish
Tissue:	Fin
Morphology:	Fibroblast
Disease:	Normal
Growth Properties:	Adherent
Age/Ethinicity:	Adult Year /
Derivation:	Derived from fin tissue of the adult salt water blue striped Grunt.
Virus Succeptility::	Does not support the replication of influenza A, herpes simplex, adeno, polio, dengue and eastern equine encephalitis viruses.
Biosafety:	2
Culture Medium:	EMEM (HBSS) (Sigma cat no. M5775) + 10mM Hepes (pH 7.3) + 2mM Glutamine +0.23% (w/v) NaHCO3 + 20% FBS (Heat Inactivated)+ 58mM NaCl
Subculturing:	Split sub-confluent cultures (70-80%) 1:2 to 1:4 using 0.05% trypsin or trypsin/EDTA; 5% CO_2; 20°C. After resuscitation split cells 1:2 to 1:4 and culture for at least one week with 1-2 media changes before shipping. Plating efficiency can be very low after resuscitation and cells may take up to 3 weeks until growth is fully established. Subculture at 80-100% confluency, not below. Cells are sensitive to plastic surface; Falcon / Corning plates and flasks have been used successfully. Fish cell lines detach easily during transit if the culture is too young.

Subculturing Medium Renewal:	1 to 2 times per week
Subculturing Subcultivation Ratio:	1:2 to 1:4
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5%
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)
Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).
References:	Proc Soc Exp Biol Med 1961;108:762; Ann NY Acad Sci 1965;126;343
Depositors:	Lanagro

Cellosaurus: CVCL_8875