

## Banco de Células do Rio de Janeiro

### **Data Sheet**

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**BCRJ Code:** 0310

**Cell Line:** HGC-27

**Species:** Homo sapiens

**Vulgar Name:** Human

Tissue: Stomach

**Cell Type:** Epithelial; Polygonal Or Short Spindle-Shaped

Morphology: **Epithelial** 

Disease: Gastric Carcinoma

**Growth Properties:** Adherent

This cell line was established by culture of the metastatic lymph node **Derivation:** from a gastric cancer patient diagnosed histological as undifferentiated

carcinoma.

**Tumor Formation::** YES

**Biosafety:** 1

Dulbecco's Modified Eagle's Medium (DMEM) with 2 mM L-glutamine, 1.0 **Culture Medium:** 

g/L glucose and 10% of fetal bovine serum.

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#### **Subculturing:**

Remove medium and rinse the adherent cells using PBS without calcium and magnesium (3-5 ml PBS for T25, 5-10ml for T75 cell culture flasks). Add Trypsin-EDTA (1-2ml per T25, 2.5ml per T75 cell culture flask), the cell sheet must be covered completely. Incubate at 37°C for 10 minutes. Carefully resuspend the cells, the addition of medium is optional but not necessary, and dispense into new flasks which contain fresh medium. Doubling Time: 17 hrs NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.

# **Subculturing Medium**

Renewal:

Every 2 to 3 days

#### **Culture Conditions:**

Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C

#### **Cryopreservation:**

95% FBS + 5% DMSO (Dimethyl sulfoxide)





**Thawing Frozen Cells:** 

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SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

- 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.
- 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes.
- 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).
- 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

AKAGI, T, KIMOTO T. HUMAN CELL LINE (HGC-27) DERIVED FROM METASTATIC LYMPH NODE OF GASTRIC CANCER. ACTA MED OKAYAMA 30(3): 215-219, 1974

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**References:** 



