

## Data Sheet

PAGE 1/5

<b>BCRJ Code:</b>	0104
<b>Cell Line:</b>	HL-60
<b>Species:</b>	Homo sapiens
<b>Vulgar Name:</b>	Human
<b>Tissue:</b>	Peripheral Blood
<b>Cell Type:</b>	Promyeloblast
<b>Morphology:</b>	Myeloblast
<b>Disease:</b>	Acute Promyelocytic Leukemia
<b>Growth Properties:</b>	Suspension
<b>Sex:</b>	Female
<b>Age/Ethnicity:</b>	36 Year / Caucasian
<b>Applications:</b>	This cell line is a suitable transfection host.
<b>DNA Profile:</b>	Amelogenin: X D5S818: 12 D13S317: 8,11 D7S820: 11,12 D16S539: 11 vWA: 16 THO1: 7,8 TPOX: 8,11 CSF1PO: 13,14
<b>Tumor Formation::</b>	Yes, in nude mice (subcutaneous myeloid tumors) Yes, in semi-solid media
<b>Products:</b>	tumor necrosis factor (TNF), also known as tumor necrosis factor alpha (TNF-alpha, TNF alpha), after stimulation with phorbol myristic acid
<b>Biosafety:</b>	1

## Data Sheet

PAGE 2/5

<b>Additional Info:</b>	HL-60 cells spontaneously differentiate and differentiation can be stimulated by butyrate, hypoxanthine, phorbol myristic acid (PMA, TPA), dimethylsulfoxide (DMSO, 1% to 1.5%), actinomycin D, and retinoic acid. The cells exhibit phagocytic activity and responsiveness to chemotactic stimuli. The line is positive for myc oncogene expression.
<b>Culture Medium:</b>	Iscove's Modified Dulbecco's Medium (IMDM) contains 4 mM L-glutamine, 4500 mg/L glucose and 20% of fetal bovine serum.
<b>Subculturing:</b>	Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at $1 \times 10^5$ viable cells/mL. Maintain cultures at a cell concentration between $1 \times 10^5$ and $1 \times 10^6$ cells/mL. NOTE: Do not allow the cell concentration to exceed $1 \times 10^6$ cells/mL. Population Doubling Time about: 24-30 hours
<b>Subculturing Medium Renewal:</b>	Every 2 to 3 days
<b>Culture Conditions:</b>	Atmosphere: air, 95%; carbon dioxide (CO <sub>2</sub> ), 5% Temperature: 37°C
<b>Cryopreservation:</b>	95% FBS + 5% DMSO (Dimethyl sulfoxide)

**Thawing Frozen Cells:**

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes.
4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio).
5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line).

**NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

**References:**

Gallagher R, et al. Characterization of the continuous, differentiating myeloid cell line (HL-60) from a patient with acute promyelocytic leukemia. *Blood* 54: 713-733, 1979. PubMed: 288488 Collins SJ, et al. Terminal differentiation of human promyelocytic leukemia cells induced by dimethyl sulfoxide and other polar compounds. *Proc. Natl. Acad. Sci. USA* 75: 2458-2462, 1978. PubMed: 276884 Collins SJ, et al. Continuous growth and differentiation of human myeloid leukaemic cells in suspension culture. *Nature* 270: 347-349, 1977. PubMed: 271272 Aggarwal BB, et al. Human tumor necrosis factor. Production, purification, and characterization. *J. Biol. Chem.* 260: 2345-2354, 1985. PubMed: 3871770 Nahm MH, et al. Identification of cross-reactive antibodies with low opsonophagocytic activity for *Streptococcus pneumoniae*. *J. Infect. Dis.* 176: 698-703, 1997. PubMed: 9291318 Berninghausen O, Leippe M. Necrosis versus apoptosis as the mechanism of target cell death induced by *Entamoeba histolytica*. *Infect. Immun.* 65: 3615-3621, 1997. PubMed: 9284127 Aparicio CL, et al. Correction for label leakage in fluorimetric assays of cell adhesion. *BioTechniques* 23: 1056-1060, 1997. PubMed: 9421636 Mansat V, et al. The protein kinase C activators phorbol esters and phosphatidylserine inhibit neutral sphingomyelinase activation, ceramide generation, and apoptosis triggered by daunorubicin. *Cancer Res.* 57: 5300-5304, 1997. PubMed: 9393753 Cuthbert JA, Lipsky PE. Regulation of proliferation and Ras localization in transformed cells by products of mevalonate metabolism. *Cancer Res.* 57: 3498-3504, 1997. PubMed: 9270019 Michael JM, et al. Resistance to radiation-induced apoptosis in Burkitt's lymphoma cells is associated with defective ceramide signaling. *Cancer Res.* 57: 3600-3605, 1997. PubMed: 9270034 Clark RA, et al. Tenascin supports lymphocyte rolling. *J. Cell Biol.* 137: 755-765, 1997. PubMed: 9151679 Tiffany HL, et al. Enhanced expression of the eosinophil-derived neurotoxin ribonuclease (RNS2) gene requires interaction between the promoter and intron. *J. Biol. Chem.* 271: 12387-12393, 1996. PubMed: 8647842 Chan YJ, et al. Synergistic interactions between overlapping binding sites for the serum response factor and ELK-1 proteins mediate both basal enhancement and phorbol ester responsiveness of primate cytomegalovirus. *J. Virol.* 70: 8590-8605, 1996. PubMed: 8970984 Mao M, et al. RIG-E, a human homolog of the murine Ly-6 family, is induced by retinoic acid during the differentiation of acute promyelocytic leukemia cell. *Proc. Natl. Acad. Sci. USA* 93: 5910-5914, 1996. PubMed: 8650192 Lepley RA, et al. Tyrosine kinase activity modulates catalysis and translocation of cellular 5-lipoxygenase. *J. Biol. Chem.* 271: 6179-6184, 1996. PubMed: 8626407 Chen H, et al. Octamer binding factors and their coactivator can activate the murine PU.1 (spi-1) promoter. *J. Biol. Chem.* 271: 15743-15752, 1996. PubMed: 8663022 U.S. Pharmacopeia USP Monographs: Technetium 99mTc Fanolesomab Injection. Rockville, MD: USP32-NF27, 2005

**Depositors:**

Ellen Jessouroun, Fundação Oswaldo Cruz

**ATCC:**

CCL-240



**Cellosaurus:** [CVCL\\_0002](#)