

Banco de Células do Rio de Janeiro

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BCRJ Code: 0105

Cell Line: HNK-1 [HNK1, Leu7]

Species: Mus musculus (B cell); Mus musculus (myeloma), mouse (B cell); mouse

(myeloma)

Vulgar Name: Mouse

Tissue: Spleen

Cell Type: Hybridoma: B Lymphocyte

Morphology: Lymphoblast

Growth Properties: Suspension

Derivation: Spleen cells were fused with P3X63Ag8.653 myeloma cells.

Products: immunoglobulin; monoclonal antibody; against human natural killer (NK) cells

and antigen dependent killer (K) cells (CD57)

Biosafety: 1

Addtional Info:

Animals were immunized with a membrane extract of the human

lymphoblastoid cell line HSB-2. Spleen cells were fused with P3X63Ag8.653 myeloma cells. The antibody also reacts with glycoproteins present on Schwann cells, oligodendrocytes and embryonic neurons. The cells will not

grow if the medium lacks 2-mercaptoethanol.

Culture Medium:

RPMI 1640 medium with 2 mM L-glutamine, 4.5 g/L glucose, 0.02 mM 2-

mercaptoethanol and 20% of fetal bovine serum.

Cultures can be maintained by addition of fresh medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension at 1 x 10e5

Subculturing: viable cells/mL. Maintain cultures at a cell concentration between 1 x 10e5 and 1 x 10e6 cells/mL. NOTE: Do not allow the cell concentration to exceed 1 x 10e6

cells/mL. Population Doubling Time about: 24-30 hours







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Subculturing Medium

Renewal:

Every 2 to 3 days

Culture Conditions:

Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C

Cryopreservation:

95% FBS + 5% DMSO (Dimethyl sulfoxide)

SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

- 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.

Thawing Frozen Cells:

- 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 x g for 5 to 7 minutes.
- 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).
- 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).





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References:

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