

**Data Sheet**

<b>BCRJ Code:</b>	0347
<b>Cell Line:</b>	JAWSII
<b>Species:</b>	Mus musculus
<b>Vulgar Name:</b>	Mouse;C57Bl/6
<b>Tissue:</b>	Bone Marrow
<b>Cell Type:</b>	Immature Dendritic Cell; Monocyte
<b>Morphology:</b>	Monocyte
<b>Growth Properties:</b>	Mixed, Adherent And Suspension
<b>Biosafety:</b>	1
<b>Culture Medium:</b>	Alpha minimum essential medium with ribonucleosides, deoxyribonucleosides, 2 mM L-glutamine, 5 ng/ml murine GM-CSF and 10% of fetal bovine serum.
<b>Subculturing:</b>	Cultures can be maintained by transferring floating cells to a centrifuge tube. Attached cells may be subcultured using 0.25% trypsin-0.03% EDTA. Pool cells and centrifuge the cell suspension at 1000 rpm for 10 minutes, resuspend the pellet in fresh medium, aspirate and dispense into new flasks. Note: This cell line grows very slowly. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.
<b>Subculturing Medium Renewal:</b>	Once a week
<b>Subculturing Subcultivation Ratio:</b>	1:2 is recommended
<b>Culture Conditions:</b>	Atmosphere: air, 95%; carbon dioxide (CO <sub>2</sub> ), 5% Temperature: 37°C

**Cryopreservation:** 95% FBS + 5% DMSO (Dimethyl sulfoxide)

**Thawing Frozen Cells:**

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). **NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

**References:**

MacKay VL, Moore EE. Immortalized dendritic cells. US Patent 5,648,219 dated Jul 15 1997 Moore EE. Preparation of immortalized cells. US Patent 5,830,682 dated Nov 3 1998

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**ATCC:**

CRL-11904