Data Sheet

BCRJ Code:	0347
Cell Line:	JAWSII
Species:	Mus musculus
Vulgar Name:	Mouse;C57Bl/6
Tissue:	Bone Marrow
Cell Type:	Immature Dendritic Cell; Monocyte
Morphology:	Monocyte
Growth Properties:	Mixed, Adherent And Suspension
Biosafety:	1
Culture Medium:	Alpha minimum essential medium with ribonucleosides, deoxyribonucleosides, 2 mM L-glutamine, 5 ng/ml murine GM-CSF and 10% of fetal bovine serum.
Subculturing:	Cultures can be maintained by transferring floating cells to a centrifuge tube. Attached cells may be subcultured using 0.25% trypsin-0.03% EDTA. Pool cells and centrifuge the cell suspension at 1000 rpm for 10 minutes, resuspend the pellet in fresh medium, aspirate and dispense into new flasks. Note: This cell line grows very slowly. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.
Subculturing Medium Renewal:	Once a week
Subculturing Subcultivation Ratio:	1:2 is recommended
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO_2), 5% Temperature: 37°C

Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)
Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).
References:	MacKay VL, Moore EE. Immortalized dendritic cells. US Patent 5,648,219 dated Jul 15 1997 Moore EE. Preparation of immortalized cells. US Patent 5,830,682 dated Nov 3 1998
Depositors:	Luiz Carlos Rodrigues Junior - Centro Universitário Fransciscano
Cellosaurus:	CVCL_3727