

## Data Sheet

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<b>BCRJ Code:</b>	0127
<b>Cell Line:</b>	K-562 LUCENA
<b>Species:</b>	Homo sapiens
<b>Vulgar Name:</b>	Human
<b>Tissue:</b>	Hematopoietic
<b>Morphology:</b>	Lymphoblast
<b>Disease:</b>	Chronic Myelogenous Leukemia
<b>Growth Properties:</b>	Suspension
<b>Sex:</b>	Female
<b>Age/Ethnicity:</b>	53 Year /
<b>Derivation:</b>	The K562-Lucena cell line was established from K562 cell line under pressure of gradual vincristine supplement in culture medium. It express the P-Glicoprotei and has a Multi Drug Resistance (MDR) phenotype.
<b>DNA Profile:</b>	Amelogenin: X CSF1PO: 9,10 D13S317: 8 D16S539: 11,12 D5S818: 11,12 D7S820: 9,11 THO1: 9.3 TPOX: 8,9 vWA: 16
<b>Tumor Formation::</b>	Yes, in nude mice Tumors developed within 21 days at 100% frequency (5/5) in nude mice inoculated subcutaneously with 107 cells.
<b>Products:</b>	Glicoprotein - P ( MDR-1)
<b>Biosafety:</b>	1
<b>Culture Medium:</b>	RPMI-1640 medium modified to contain 2 mM L-glutamine, 4500 mg/L glucose, 60nM vincristine and 10% of fetal bovine serum.

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### Subculturing:

Cultures can be maintained by the addition or replacement of fresh medium. Start new cultures at  $1 \times 10^5$  viable cells/mL. Subculture at  $1 \times 10^6$  cells/mL. T-75 flasks are recommended for subculturing this product.

### Subculturing Medium Renewal:

Every 2 to 3 days

### Culture Conditions:

Atmosphere: air, 95%; carbon dioxide (CO<sub>2</sub>), 5% Temperature: 37°C

### Cryopreservation:

95% FBS + 5% DMSO (Dimethyl sulfoxide)

### Thawing Frozen Cells:

**SAFETY PRECAUTION:** It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately  $125 \times g$  for 5 to 7 minutes. 4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line).  
**NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

### References:

Ciência e Cultura, 46:63-69, 1994 Braz.J. Med. Biol. Res, 29:401-542, 1996

### Depositors:

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### Cellosaurus:

[CVCL\\_D162](#)



