Data Sheet

BCRJ Code:	0138
Cell Line:	L1210
Species:	Mus musculus
Vulgar Name:	Mouse; Dba Subline 212
Tissue:	Skin
Morphology:	Lymphoblast
Disease:	Lymphocytic Leukemia
Growth Properties:	Suspension
Sex:	Female
Age/Ethinicity:	8 Month /
Derivation:	established from the methylcholanthrene-induced tumor in a DBA strain mouse (subline 212, 8-month-old female) in 1949; the cells were propagated by heterotransplantation into DBA/2 mice; an in-vitro suspension culture was first reported in 1966
Applications:	cells are used for cytotoxicity studies
Tumor Formation::	Yes, in nude mice
Biosafety:	1
Culture Medium:	Dulbecco's modified Eagle's medium with 2 mM L-glutamine, 4.5 g/L glucose and 10% of horse serum.

Subculturing:	Cultures can be maintained by the addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension. Start cultures at 5 x 10 4 viable cells/mL. Population Doubling Time: 8 to 10 hrs
Subculturing Medium Renewal:	Add fresh medium (20% to 30% by volume) every 2 to 4 days.
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)
Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

References:	J Nat Cancer Inst 1949; 10:179; J Nat Cancer Inst 1966;36:405 Law LW, et al. Observations on the effect of a folic-acid antagonist on transplantable lymphoid leukemias in mice. J. Natl. Cancer Inst. 10: 179-192, 1949. PubMed: 18143260 Moore GE, et al. Suspension Cell Culture and In Vivo and In Vitro Chromosome Constitution of Mouse Leukemia L1210. J. Natl. Cancer Inst. 36: 405-421, 1966. PubMed: 18630317 Himmelfarb P, et al. Growth of Colonies of. Murine Leukemia L1210 in Vitro. Cancer Chemother. Rep. 51: 451-3 1967 Young SW, et al. Gadolinium(III) texaphyrin: a tumor selective radiation sensitizer that is detectable by MRI. Proc. Natl. Acad. Sci. USA 93: 6610-6615, 1996. PubMed: 8692865
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