

Data Sheet

PAGE 1/4

BCRJ Code:	0165
Cell Line:	MDA-MB-435S
Species:	Homo sapiens
Vulgar Name:	Human
Tissue:	Previously Described As: Mammary Gland/Breast; Derived From Metastatic Site: Pleural Effusion
Cell Type:	Melanocyte,Melanoma
Morphology:	Spindle Shaped
Disease:	Previously Described As Ductal Carcinoma
Growth Properties:	Adherent
Sex:	Female
Age/Ethnicity:	31 Year / Caucasian
Derivation:	MDA-MB-435S is a spindle shaped strain which evolved from the parent line (MDA-MB-435) isolated in 1976 (R. Cailleau et al.) from the pleural effusion of a 31 year old female with metastatic ductal adenocarcinoma of the breast. Parental cells displayed the dispersed pattern (type II) when treated with immunofluorescent stain for tubulin. However, recent studies have generated questions about the origin of the parent cell line, MDA-MB-435, and by extension MDA-MB-435S. Gene expression analysis of the cells produced microarrays in which MDA-MB-435 clustered with cell lines of melanoma origin instead of breast [PubMed ID: 10700174, PubMed ID: 15150101, PubMed ID: 15679052].
DNA Profile:	Amelogenin: X CSF1PO: 11 D13S317: 12 D16S539: 13 D5S818: 12 D7S820: 8,10 THO1: 6,7 TPOX: 8,11 vWA: 16,18
Tumor Formation::	No, in immunosuppressed mice Yes, in semisolid medium

Data Sheet

PAGE 2/4

Products:	Tubulin, actin
Biosafety:	1
Culture Medium:	Leibovitz's L-15 Medium contains 2 mM L-glutamine, NO sodium bicarbonate and fetal bovine serum to a final concentration of 10%. Note: The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air. A CO ₂ and air mixture is detrimental to cells when using this medium for cultivation.
Subculturing:	Remove and discard culture medium. Briefly rinse the cell layer with PBS without calcium and magnesium to remove all traces of serum which contains trypsin inhibitor. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.
Subculturing Medium Renewal:	2 to 3 times per week
Subculturing Subcultivation Ratio:	1:3 to 1:6
Culture Conditions:	Atmosphere: air, 100% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)

Thawing Frozen Cells:

SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris.

1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions.
3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes.
4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio).
5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line).

NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).

Data Sheet

PAGE 4/4

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Cellosaurus:

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