

Data Sheet

BCRJ Code:	0165
Cell Line:	MDA-MB-435S
Species:	Homo sapiens
Vulgar Name:	Human
Tissue:	Previously Described As: Mammary Gland/Breast; Derived From Metastatic Site: Pleural Effusion
Cell Type:	Melanocyte, Melanoma
Morphology:	Spindle Shaped
Disease:	Previously Described As Ductal Carcinoma
Growth Properties:	Adherent
Sex:	Female
Age/Ethnicity:	31 Year / Caucasian
Derivation:	MDA-MB-435S is a spindle shaped strain which evolved from the parent line (MDA-MB-435) isolated in 1976 (R. Cailleau et al.) from the pleural effusion of a 31 year old female with metastatic ductal adenocarcinoma of the breast. Parental cells displayed the dispersed pattern (type II) when treated with immunofluorescent stain for tubulin. However, recent studies have generated questions about the origin of the parent cell line, MDA-MB-435, and by extension MDA-MB-435S. Gene expression analysis of the cells produced microarrays in which MDA-MB-435 clustered with cell lines of melanoma origin instead of breast [PubMed ID: 10700174, PubMed ID: 15150101, PubMed ID: 15679052].
DNA Profile:	Amelogenin: X CSF1PO: 11 D13S317: 12 D16S539: 13 D5S818: 12 D7S820: 8,10 THO1: 6,7 TPOX: 8,11 vWA: 16,18
Tumor Formation::	No, in immunosuppressed mice Yes, in semisolid medium

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Products: Tubulin, actin

Biosafety: 1

Culture Medium: Leibovitz's L-15 Medium contains 2 mM L-glutamine, NO sodium bicarbonate and fetal bovine serum to a final concentration of 10%. Note: The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air. A CO₂ and air mixture is detrimental to cells when using this medium for cultivation.

Subculturing: Remove and discard culture medium. Briefly rinse the cell layer with PBS without calcium and magnesium to remove all traces of serum which contains trypsin inhibitor. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.

Subculturing Medium Renewal: 2 to 3 times per week

Subculturing Subcultivation Ratio: 1:3 to 1:6

Culture Conditions: Atmosphere: air, 100% Temperature: 37°C

Cryopreservation: 95% FBS + 5% DMSO (Dimethyl sulfoxide)

Thawing Frozen Cells:

SAFETY PRECAUTION: It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes.
4. Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio).
5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line).

NOTE: It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

References:

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Chambers AF. MDA-MB-435 and M14 cell lines: identical but not M14 melanoma? *Cancer Res.* 69(13): 5292-5293, 2009. PubMed: 19549886.

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ATCC:

HTB-129

Cellosaurus:

[CVCL_0622](https://www.ebi.ac.uk/ces/entry/CVCL_0622)