# Data Sheet



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| **BCRJ Code:** | 0262 |
| **Cell Line:** | MEG-01 |
| **Species:** | Homo sapiens |
| **Vulgar Name:** | Human |
| **Tissue:** | Bone Marrow |
| **Cell Type:** | Megakaryoblast |
| **Morphology:** | Lymphoblast |
| **Disease:** | Chronic Myelogenous Leukemia, |
| **Growth Properties:** | Mixed, Adherent And Suspension |
| **Sex:** | Male |
| **Age/Ethinicity:** | 55 Year / |
| **Derivation:** | The MEG-01 cell line was derived in 1983 at the Nagoya University School of Medicine, Nagoya, Japan from bone marrow cells taken from a patient in megakaryoblastic crisis of CML. |
| **Applications:** | This cell line is a suitable transfection host. |
| **DNA Profile:** | Amelogenin: X,Y CSF1PO: 10 D13S317: 8 D16S539: 9 D5S818: 13 D7S820: 11 THO1: 7 TPOX: 8,11 vWA: 16 |
| **Biosafety:** | 1 |
| **Culture Medium:** | RPMI-1640 medium modified to contain 2 mM L-glutamine, 4500 mg/L glucose and 10% of fetal bovine serum. |

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| **Subculturing:** | Subcultures are prepared by scraping the adherent cells into the medium. From the resulting suspension dilute cells to a concentration 1-2 X 10e5 cells/mL into fresh medium in a new flasks. Keep culture below approximate density of 10e6 cells/mL. Subcultures are prepared by scraping the adherent cells into the medium, and diluting the resulting suspension into fresh medium in new flasks. |
| **Subculturing Medium Renewal:** | 2 to 3 times per week |
| **Culture Conditions:** | Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C |
| **Cryopreservation:** | 95% FBS + 5% DMSO (Dimethyl sulfoxide) |
| **Thawing Frozen Cells:** | SAFETY PRECAUTION: Is highly recommend that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the Oring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4.Discard the supernatant and Resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in a appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6). |
| **References:** | 23348: Ogura M, et al. Establishment of a novel human megakaryoblastic leukemia cell line, MEG- 01, with positive Philadelphia chromosome. Blood 66: 1384-1392, 1985. PubMed: 2998511 |

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| **ATCC:** | CRL-2021 |