

Data Sheet

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BCRJ Code:	0382
Cell Line:	SW 1353
Species:	Homo sapiens
Vulgar Name:	Human
Tissue:	Bone
Morphology:	Fibroblast
Disease:	Chondrosarcoma
Growth Properties:	Adherent
Sex:	Female
Age/Ethnicity:	72 Year / Caucasian
Derivation:	The SW 1353 cell line was initiated by A. Leibovitz at the Scott and White Clinic, Temple, Texas in 1977 from a primary grade II chondrosarcoma of the right humerus obtained from a 72 year old female Caucasian.
Applications:	Transfection host
Biosafety:	1
Culture Medium:	Leibovitz's L-15 Medium and fetal bovine serum to a final concentration of 10%. The L-15 medium formulation was devised for use in a free gas exchange with atmospheric air.
Subculturing:	Remove medium, and rinse with 0.25% trypsin, 0.03% EDTA solution. Remove the solution and add an additional 1 to 2 mL of trypsin-EDTA solution. Allow the flask to sit at room temperature (or at 37°C) until the cells detach. Add fresh culture medium, aspirate and dispense into new culture flasks.

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Subculturing Medium Renewal:

2 to 3 times per week

Subculturing Subcultivation Ratio:

1:02

Culture Conditions:

Atmosphere: air, 100%; carbon dioxide (CO₂), 0%

Cryopreservation:

95% FBS + 5% DMSO (Dimethyl sulfoxide)

Thawing Frozen Cells:

SAFETY PRECAUTION: It is highly recommended that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submerged in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris. 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions. 3. For cells that are sensitive to DMSO it is recommended that the cryoprotective agent be removed immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125 x g for 5 to 7 minutes. 4. Discard the supernatant and resuspend cell pellet with the recommended complete medium (see the specific batch information for the culture recommended dilution ratio). 5. Incubate the culture in an appropriate atmosphere and temperature (see "Culture Conditions" for this cell line). **NOTE:** It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

Depositors:

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ATCC:

HTB-94