Data Sheet

BCRJ Code:	0235
Cell Line:	Toledo
Species:	Homo sapiens
Vulgar Name:	Human
Tissue:	Peripheral Blood
Cell Type:	B Lymphocyte
Morphology:	Lymphoblast
Disease:	Diffuse Large Cell Lymphoma; Non-Hodgkin'S B Cell Lymphoma
Growth Properties:	Suspension
Sex:	Female
Age/Ethinicity:	ADULT / White
Derivation:	Toledo was established in 1990 from peripheral blood leukocytes (PBL) of a patient that originally had a diffuse large cell lymphoma (DLCL). [53289]
Applications:	This cell line is a model system for studying non-Hodgkin lymphomas.
DNA Profile:	D5S818:10,12 D7S820: 9,10 THO1: 8,9.3 TPOX: 8,11 vWA: 16,17 Amelogenin: X CSF1PO: 12 D13S317:11 D16S539:9
Biosafety:	1

Addtional Info:	Following high-dose chemotherapy and bone marrow transplantation, the patient subsequently developed a lymphoma in the brain. Though the morphology of the cells resembles Burkitt's lymphoma, the cells lack the typical chromosomal translocations of Burkitt's lymphoma. The karyotype does exhibit multiple chromosomal aberrations. The cells do not express surface or cytoplasmic immunoglobulin.
Culture Medium:	RPMI-1640 medium modified to contain 2 mM L-glutamine, 4500 mg/L glucose and 10% of fetal bovine serum.
Subculturing:	Cultures can be maintained by addition of fresh medium or replacement of medium. Alternatively, cultures can be established by centrifugation with subsequent resuspension in fresh medium at 3 to 5 x 10e5 viable cells/mL. Maintain cultures at cell concentrations between 3 x 10e5 and 3 x 10e6 viable cells/mL. Population Doubling Time: 24 to 30 hrs
Subculturing Medium Renewal:	Every 2 to 3 days
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)

Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).
References:	53289: Gabay C, et al. Somatic mutations and intraclonal variations in the rearranged Vkappa genes of B-non-Hodgkin's lymphoma cell lines. Eur. J. Immunol. 63: 180-191, 1999. PubMed: 10485273
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