BCRJ Code:	0236
Cell Line:	TOV-21G
Species:	Homo sapiens
Vulgar Name:	Human
Tissue:	Ovary
Morphology:	Epithelial
Disease:	Primary Malignant Adenocarcinoma
Growth Properties:	Adherent
Sex:	Female
Age/Ethinicity:	62 Year /
Derivation:	This cell line was initiated in October of 1991 from a patient of French-Canadian descent with no family history of ovarian cancer. [49408]
DNA Profile:	Amelogenin: X CSF1PO: 13,15 D13S317: 11,12 D16S539: 10,12 D5S818: 12,13 D7S820: 12 THO1: 7,9.3 TPOX: 8,11 vWA: 17
Tumor Formation::	Yes, the cells are tumorigenic in nude mice
Products:	keratin
Biosafety:	1
Culture Medium:	1:1 mixture of MCDB 105 medium and Medium 199 with fetal bovine serum to a final concentration of 15%.

Subculturing:	Volumes used in this protocol are for 75 cm2 flask; proportionally reduce or increase amount of dissociation medium for culture vessels of other sizes. Remove and discard culture medium. Briefly rinse the cell layer with PBS without calcium and magnesium to remove all traces of serum which contains trypsin inhibitor. Add 2.0 to 3.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C. NOTE: For more information on enzymatic dissociation and subculturing of cell lines consult Chapter 12 in Culture of Animal Cells, a manual of Basic Technique by R. Ian Freshney, 6th edition, published by Alan R. Liss, N.Y., 2010.
Subculturing Medium Renewal:	Every 3 to 4 days
Subculturing Subcultivation Ratio:	1:3 to 1:4
Culture Conditions:	Atmosphere: air, 95%; carbon dioxide (CO2), 5% Temperature: 37°C
Cryopreservation:	95% FBS + 5% DMSO (Dimethyl sulfoxide)

Thawing Frozen Cells:	SAFETY PRECAUTION: It is strongly recommended to always wear protective gloves, clothing, and a full-face mask when handling frozen vials. Some vials may leak when submerged in liquid nitrogen, allowing nitrogen to slowly enter the vial. Upon thawing, the conversion of liquid nitrogen back to its gas phase may cause the vial to explode or eject its cap with significant force, creating flying debris. 1. Thaw the vial by gently agitating it in a 37°C water bath. To minimize contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes). 2. Remove the vial from the water bath as soon as its contents are thawed and decontaminate it by dipping in or spraying with 70% ethanol. From this point, all operations must be performed under strict aseptic conditions. 3. For cells sensitive to DMSO, it is recommended to remove the cryoprotective agent immediately. Transfer the vial contents to a centrifuge tube containing 9.0 mL of complete culture medium and centrifuge at approximately 125 × g for 5 to 7 minutes. 4. Discard the supernatant and resuspend the cell pellet in the recommended complete medium (see specific batch information for the appropriate dilution ratio). 5. Incubate the culture under appropriate atmospheric and temperature conditions (see "Culture Conditions" for this cell line). NOTE: It is important to avoid excessive alkalinity of the medium during cell recovery. To minimize this risk, it is recommended to place the culture vessel containing the growth medium in the incubator for at least 15 minutes before adding the vial contents. This allows the medium to stabilize at its normal pH (7.0 to 7.6).
References:	42090: Mes-Masson AM, Provencher D. Primary cultures of normal and tumoral human ovarian epithelium. US Patent 5,710,038 dated Jan 20 1998 49408: Provencher DM, et al. Characterization of four novel epithelial cancer cell lines. In Vitro Cell. Dev. Biol.
Depositors:	CYNTIA OLIVEIRA; INCA
Cellosaurus:	CVCL_3613